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Title	Micellization of dibucaine hydrochloride (DC) in water by measurement of the fluorescence of DC
Sub Title	
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Publisher	共立薬科大学
Publication year	1995
Jtitle	共立薬科大学研究年報 (The annual report of the Kyoritsu College of
	Pharmacy). No.40 (1995.) ,p.1- 4
JaLC DOI	
Abstract	
Notes	
Genre	Technical Report
URL	https://koara.lib.keio.ac.jp/xoonips/modules/xoonips/detail.php?koara_id=AN00062898-00000040-0001

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共立薬科大学 Ann. Rept. Kyoritsu Coll. Pharm.

Micellization of Dibucaine Hydrochloride (DC) in Water by Measurement of the Fluorescence of DC

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The micellization of dibucaine hydrochloride (DC) in water at $25\,^{\circ}$ C was investigated by measuring the fluorescence spectrum of DC. Furthermore, the surface tension of an aqueous solution of DC was measured with a Du Nöuy tensiometer at $25\,^{\circ}$ C. The critical micelle concentration of DC in water at $25\,^{\circ}$ C was estimated as $7\,^{\circ}$ 8 × 10^{-2} M by the fluorescence method, and this value is coincided with the value of $7.4\,^{\circ}$ 10⁻² M by the surface tension method.

Key words dibucaine hydrochloride; micelle; fluorescence spectrum; critical micelle concentration; surface tension

Dibucaine hydrochloride (DC) is a local anesthetic drug. It is important to study the surface activity of DC in connection with its pharmacological effects at the surface of a biological membrane since the parallelism between the surface activity and the pharmacological potency was proposed by Levy¹. The aggregation of DC in water has been studied by the light scattering method² and the drop volume method³.

On the other hand, ammonium 8-anilino-1-naphthalene-sulfonate (ANS) as a fluorescent probe has been used for the measurements of critical micelle concentration $(cmc)^4$, critical reversed micelle concentration⁵⁾ and the surface potential of the micelles⁶⁾. This is based on the fact that ANS adsorbed on the micelle surface fluorescess strongly although the fluorescence of free ANS is very weak. Recently, the study on the pK_a of DC by measuring the fluorescence of DC has been reported⁷⁾. The micellization of non-fluorescent compound can be determined by the ANS method, but the micellization of fluorescent compound can not be accurately determined by the ANS method because fluorescence intensity increases with increasing concentration of fluorescent compound. By the way, it is well known that the physicochemical properties of surfactant are largely changed at the cmc. So the maximum wavelength of emission spectrum and the quantum yield of fluorescent compound may be changed near the cmc. The fluorescence of DC has been reported as a function of pH⁷⁾. However, the fluorescence of DC has not yet been determined by varying the concentration of DC.

From these points of view, we measured the fluorescence spectrum of aqueous solution of DC at various concentrations and estimated the cmc of DC. Furthermore, the cmc of DC obtained from the fluorescence method was compared with that obtained from the surface tension method.

Experimental

Materials Dibucaine hydrochloride (DC) purchased from Wako Pure Chemical Industries, Ltd. was recrystallized twice from methanol. The molecular structure of DC is shown in Chart 1. Deionized and twice-distilled water was used throughout this study.

Measurement of Fluorescence Spectrum DC was dissloved in water. The fluorescence spectra were measured with a Hitachi F-4000 spectrofluorometer as previously described ⁶⁾. The temperature was

$$\begin{array}{c}
N \\
O(CH_2)_3CH_3 \\
\bullet HCI \\
CONHCH_2CH_2N(C_2H_5)_2
\end{array}$$

Chart 1 Molecular Structure of Dibucaine Hydrochloride (DC)

maintained at 25 ± 0.2 °C during the fluorescence measurement by circulating water through the cuvette holders. The fluorescence spectra were determined after temperature of the sample solution had reached the measurement temperature. The wavelength of excitation was 373 nm. Louro *et al.*⁷⁾ reported that the wavelength of excitation (λ_{ex}) for the aqueous solution of DC was 328 nm. However, we obtained 373 nm of λ_{ex} when we searched the suitable λ_{ex} at a fixed wavelength of emission. Thus, we measured the fluorescence spectra of aqueous solutions of DC at excitation wavelength 373 nm.

Measurement of Surface Tension The surface tension was measured with a Du Nöuy tensiometer. The platinum ring with a diameter of 19 mm was heated by an oxidizing flame before use. The thermostat temperature was maintained at 25 ± 0.1 °C. For the calculation of the surface tension of aqueous solutions, the value of 71.96 mN m⁻¹ was used as the surface tension of pure water at 25°C. The experimental determination of the surface tension was precise to ±0.1 mN m⁻¹.

Results and Discussion

Maximum Wavelength of Emission Changes in the maximum wavelength of the emission spectrum (λ_{max}) of DC excited at 373 nm by varying the concentration of DC from 0 to 0.2 M are shown in Fig. 1, where C is the concentration of DC. DC in water had a λ_{max} at 418 nm, and λ_{max} was constant at concentrations up to 4×10^{-2} M. At 6×10^{-2} M λ_{max} shifted largely to a longer wavelength. At 8×10^{-2} M λ_{max} further shifted to a longer wavelength, and the value of λ_{max} remained constant at concentrations above 8×10^{-2} M. These phenomena are considered to be caused by that the premicellar associations begins to be formed at about 6×10^{-2} M and that the micelles are formed at concentrations above 8×10^{-2} M. In general, the changes in fluorescence followed by a premicellar association are larger as compared with the other physicochemical parameters. In addition, surfactants with aromatic hydrophobic groups associate by a process⁸ in which aggregate growth occurs by the continuous stepwise addition of monomers. The cmc of DC in water at 25°C is estimated as $7 \sim 8 \times 10^{-2}$ M. The cmc of DC in water is reported as $6.0 \sim 6.6 \times 10^{-2}$ M at 30°C by the light scattering method² and 7.87×10^{-2} M at 25°C by the drop volume technique. So the value of cmc of DC obtained by the fluorescence method is considered to by adequate.

Surface Tension of Aqueous Solution of DC The surface tension (γ) of DC in water as a function of log C was presented in Fig. 2. As can be seen in Fig. 2, γ decreased with increasing concentration of DC and became nearly constant at concentrations of DC above 8×10^{-2} M. The inflection point in the γ vs. log C curve is not sharp. This phenomenon is satisfied with the results shown in Fig. 1. This is considered to be based on the association mechanism of DC in which aggregate growth occurs by the continuous stepwise addition. The value of cmc in the γ vs. log C curve was estimated as the point of intersection of the extension of the gentle curve and horizontal line: the cmc of DC in water was obtained as 7.4×10^{-2} M. This value is coincided with the value obtained by the fluorescence method.

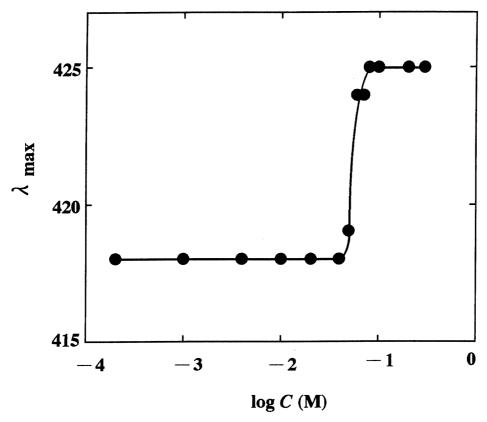


Fig. 1 Variation of λ_{max} of DC with Increasing Concentration of DC

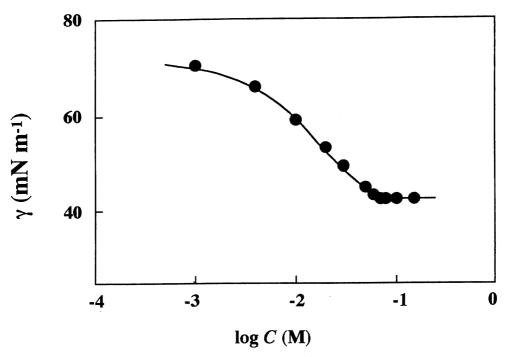


Fig. 2 Surface Tension of Aqueous Solutions of DC at 25°C

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In conclusion, the micellization of DC in water can be determined by measuring the self-fluorescence of DC. The cmc of DC in water at 25% was obtained as $7\sim8\times10^{-2}\,\mathrm{M}$ by the fluorescence method, and this value was coincided with the cmc value of $7.4\times10^{-2}\,\mathrm{M}$ obtained by the surface tension method.

References

- 1) J. V. Levy, J. Pharm. Pharmacol., 20, 813 (1968).
- 2) D. Attowood, P. Fletcher, J. Pharm. Pharmacol., 38, 494 (1986).
- 3) H. Matsuki, S. Hashimoto, S. Kaneshina, Langmuir, 10, 1882 (1994).
- 4) S. Yokoyama, Y. Fujino, M. Kondo, T. Fujie, *Chem. Pharm. Bull.*, 43, 1055 (1995); H. C. Chiang, A. Lukton, *J. Phys. Chem.*, 79, 1935 (1975); K. S. Birdi, H. N. Singh, S. V. Dalsager, *ibid.*, 83, 2733 (1979).
- 5) S. Yokoyama, T. Fujie, Chem. Pharm. Bull., 38, 2249 (1990); S. Yokoyama, Y. Fujino, T. Fujie, ibid., 41, 2026 (1993).
- 6) S. Yokoyama, A. Kaneko, T. Fujie, Bull. Chem. Soc. Jpn., 61, 3451 (1988).
- 7) S. R. W. Louro, O. R. Nascimento, M. Tabak, Biochim. Biophys. Acta. 1190, 319 (1994).
- 8) S. Yokoyama, Y. Fujino, Y. Kawamoto, A. Kaneko, T. Fujie, *Chem. Pharm. Bull.*, 42, 1351 (1994); P. Mukerjee, J. Pharm. Sci., 63, 972 (1974).