

Title	Studies of cirratiomycins. part II. the structures of cirratiomycin A and B, the new peptide antibiotics
Sub Title	
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Publisher	共立薬科大学
Publication year	1982
Jtitle	共立薬科大学研究年報 (The annual report of the Kyoritsu College of Pharmacy). No.27 (1982. ) ,p.65- 71
JaLC DOI	
Abstract	
Notes	抄録
Genre	Technical Report
URL	<a href="https://koara.lib.keio.ac.jp/xoonips/modules/xoonips/detail.php?koara_id=AN00062898-00000027-0065">https://koara.lib.keio.ac.jp/xoonips/modules/xoonips/detail.php?koara_id=AN00062898-00000027-0065</a>

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## Studies of Cirratiomycins. Part II.

### The Structures of Cirratiomycin A and B, the New Peptide Antibiotics†

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Toyoshige ENDŌ, Haruo SETO\* and Noboru ŌTAKE\*

In the preceding paper, we described the production, isolation, physico-chemical properties and biological activities of cirratiomycin A (1) and B (2) produced by *Streptomyces cirratus* 248-Sq2. These two antibiotics are very closely related to each other in structure. This report presents the structural elucidation of cirratiomycin A and B in detail.

The 400 MHz  $^1\text{H-NMR}$  spectrum of 1 (Fig. 1) revealed 40 well-resolved unexchangeable proton signals. Based on the chemical shift trends, these were classified as follows; 6 methyls, with one allylic methyl ( $\delta$  1.82 ppm); 7 methylenes, of which three were oxygenated; 7 aliphatic methines and one olefinic methine. Exhaustive spin decoupling experiments proved the partial structures shown in the next page.

The 100 MHz  $^{13}\text{C-NMR}$  spectrum of 1 (Table I) showed a total of 30 carbon signals, with one of double intensity. All protonated carbons were analyzed by selective proton decoupling experiments. The other carbons were as follows; seven carbonyl carbons

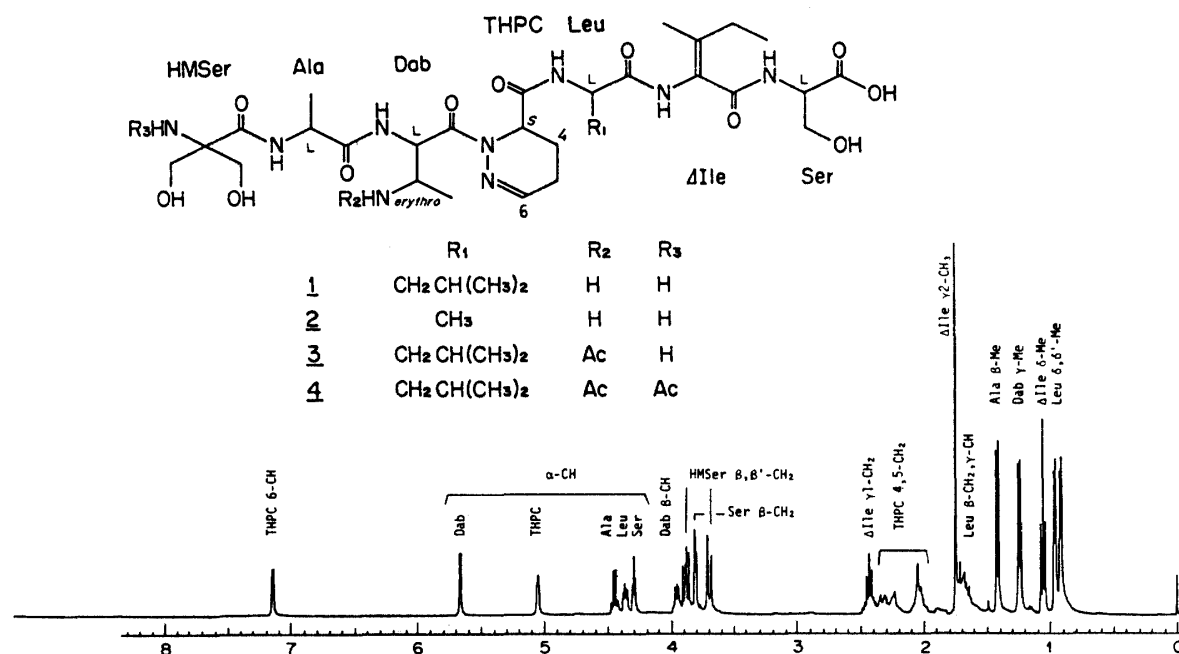


Fig. 1. 400MHz  $^1\text{H-NMR}$  Spectrum of Cirratiomycin A in  $\text{D}_2\text{O}$

† Agr. Biol. Chem. 46 (7) 1891—1898, (1982) より転載.

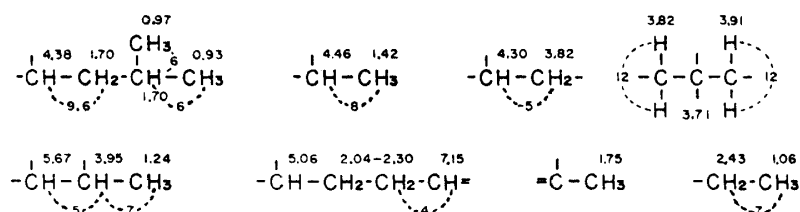
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Table I.  $^{13}\text{C}$ -Chemical Shift of Cirratiomycin A (dioxane as an internal standard, 67.4 ppm)

Amino acid		ppm	Amino acid		ppm
HMSer	$\alpha$ -C	66.2	Leu	$\alpha$ -CH	53.6
	$\beta$ 1, $\beta$ 2-CH <sub>2</sub>	62.4		$\beta$ -CH <sub>2</sub>	40.4
Ala	$\alpha$ -CH	51.0		$\gamma$ -CH	25.2
	$\beta$ -CH <sub>3</sub>	17.3		$\delta$ 1-CH <sub>3</sub>	22.8
Dab	$\alpha$ -CH	53.0	$\delta$ 2-CH <sub>3</sub>	21.7	
	$\beta$ -CH	48.8	Ile	$\alpha$ -C	122.5
	$\gamma$ -CH <sub>3</sub>	14.0		$\beta$ -C	149.3
THPC	C-3	53.7		$\gamma$ 1-CH <sub>2</sub>	27.5
	C-4	19.9*	$\gamma$ 2-CH <sub>3</sub>	18.7	
	C-5	20.6*	$\delta$ -CH <sub>3</sub>	12.9	
	C-6	149.1	Ser	$\alpha$ -CH	57.8
CO		167.7		$\beta$ -CH <sub>2</sub>	63.2
		171.3			
		174.6			
		176.7			

\* Interchangeable.

Partial structures



( $\delta$  167–177 ppm), two olefinic ( $\delta$  122, 149 ppm) and one aliphatic ( $\delta$  66.2 ppm) quaternary carbon. The existence of one C=C and one C=N bond, which are suggested by the three olefinic carbon signals, and seven C=O bonds indicate the presence of a ring structure in **1** (10-degrees of unsaturation).

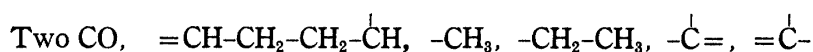
Since the IR spectrum of **1** suggested the presence of peptide bonds,  $\nu_{\text{max}}^{\text{KBr}}$  1650  $\text{cm}^{-1}$ , **1** (0.5 mg) was hydrolyzed with 6 N HCl, 120°C for 16 hours and an amino acid analysis was carried out using an amino acid autoanalyzer. As a result, one mol each of Leu, Ala and Ser were detected together with two unidentified peaks. One was observed shortly before Asp and the other shortly before  $\text{NH}_3$ .

In order to isolate these amino acids, 10 mg of **1** was hydrolyzed (6 N HCl, 120°C, 16 hours) in a sealed tube. After removal of the HCl *in vacuo*, the residue was passed through a column of Dowex 50 W-X2 (0.5 M pyridine-AcOH buffer, pH 6.4) to separate a neutral amino acid fraction and one basic amino acid which was identified as a diastereomeric mixture of  $\alpha, \beta$ -diaminobutyric acid (abbreviated as Dab) by  $^1\text{H-NMR}$ . It has been

reported that under this hydrolysis condition, racemization occurs at the  $\alpha$ -carbon atom of Dab. The different magnetic environments of the two protons,  $H\alpha$  and  $H\beta$ , between the diastereomeric isomers are reflected in the signals having different coupling constants (7 Hz *erythro* and 4 Hz *threo*). The absolute configuration of Dab in the native antibiotic will be discussed later.

Then, the neutral amino acid fraction was successively separated into four amino acids by Sephadex G-25 partition column chromatography (*n*-BuOH-AcOH-water, 4 : 1 : 2). The first three were identified as L-Leu, L-Ala and L-Ser, and the last unknown amino acid was determined as hydroxymethylserine (HMSer) by direct comparison ( $^1\text{H-NMR}$ , amino acid analysis) with an authentic sample.

In the  $^1\text{H-NMR}$  spectrum, the resonances due to these five amino acids which showed the presence in an equimolar ratio were easily assigned, while the  $^{13}\text{C-NMR}$  spectrum suggested further the existence of two acid-labile amino acids, temporarily designated as X and Y, whose partial structures were as follows ;



The  $^1\text{H-NMR}$  spectrum of **1** in  $\text{DMSO-}d_6$  (Fig. 2) afforded four doublet amide protons as well as a singlet. Spin decoupling experiments irradiating at each  $\alpha$ -methine proton permitted us to assign these doublet  $\alpha$ -amide protons as Leu, Ala, Ser and Dab ( $\delta$  8.49, 8.68, 7.30 and 8.22 ppm, respectively).

The dissociation constants,  $\text{p}K_a'$ , of **1** were determined as 3.4, 6.7 and 8.8 by potentiometric titration. These values suggested the presence of three functional groups, *i.e.* one carboxylic and two amino groups which were well reflected in *N*-acetylation experiments. Treatment of **1** (10 mg) with acetic anhydride in MeOH ( $0^\circ\text{C}$ , 30 min) gave 7 mg of *N*-monoacetyl cirratiomycin A (**3**) and 3 mg of *N*-diacetyl cirratiomycin A (**4**). Repeated acetylation of **3** under the same conditions gave **4**. In the  $^1\text{H-NMR}$  spectrum of **3** (in  $\text{D}_2\text{O}$ ), the  $\beta$ -methine proton of Dab was shifted downfield from 4.0 ppm for the parent antibiotic to 4.4 ppm, at the same time as an upfield shift of Dab  $\gamma$ -Me (from 1.24 to 1.08 ppm), indicating the  $\beta$ -amino group of Dab to be free. On the other hand, the  $^1\text{H-NMR}$  spectrum of **4** closely resembled that of **3** except for signals assignable to HM-Ser, suggesting that the second acetylation took place at a free amino group attached to the quaternary carbon. In agreement with this, a new singlet amide proton (7.70 ppm) appeared in the spectrum of **4** in  $\text{DMSO-}d_6$  (Fig. 3), together with the doublet  $\beta$ -amide proton of Dab (7.10 ppm). These results are compatible with the  $\text{p}K_a'$  values of 8.8 and 6.4 which were assigned to  $\beta\text{-NH}_2$  of Dab and  $\text{NH}_2\text{-C}\zeta$ , respectively. Treatment of **1** with dansyl chloride (DNS-Cl) at pH 9.5 only dansylated the  $\beta$ -amino group of Dab but did not give a bis DNS derivative.

Because **1** contained acid-labile amino acids, enzymatic reactions (endo and exopeptidases) were tried to obtain X and Y as intact. However, these degradations were

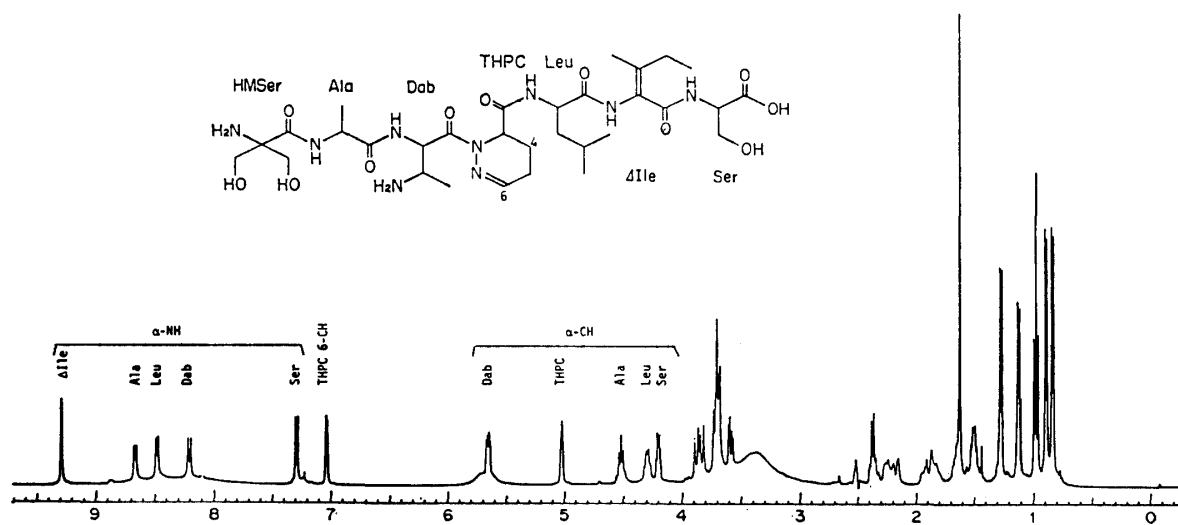


Fig. 2.  $^1\text{H-NMR}$  Spectrum of Cirratiomycin A in  $\text{DMSO-}d_6$ .

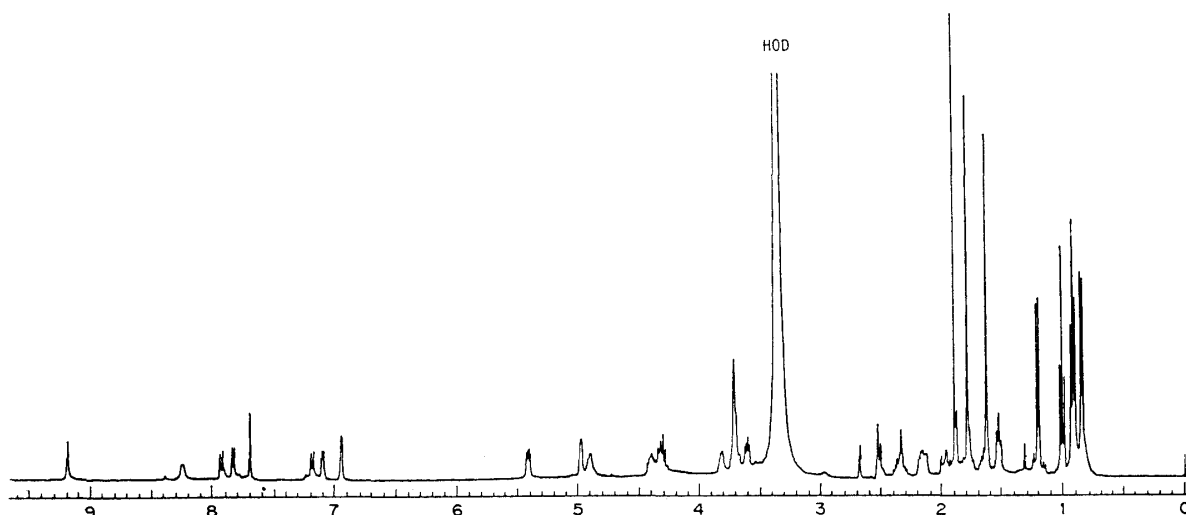


Fig. 3.  $^1\text{H-NMR}$  Spectrum of *N*-Diacetyl Cirratiomycin A in  $\text{DMSO-}d_6$ .

not successful due to the presence of unusual amino acids. Then, efforts were made to isolate small peptide fragments under several hydrolysis conditions. Mild acid hydrolysis of 1 (6 N HCl, 37°C, 7 days) and subsequent purification by column chromatography using Toyopearl HW 40 F and Dowex 50 W-X 2 (0.5 M pyridine-AcOH buffer, pH 6.4) afforded four peptide fragments. The *N*-terminal residues of these fragments were determined by the DNS method. The  $^1\text{H-NMR}$  spectra (6, 7) are shown in Fig. 4a, b.

HMSer-Ala	5
Dab-X-Leu	6
Leu-Y	7
Leu-Y-Ser	8

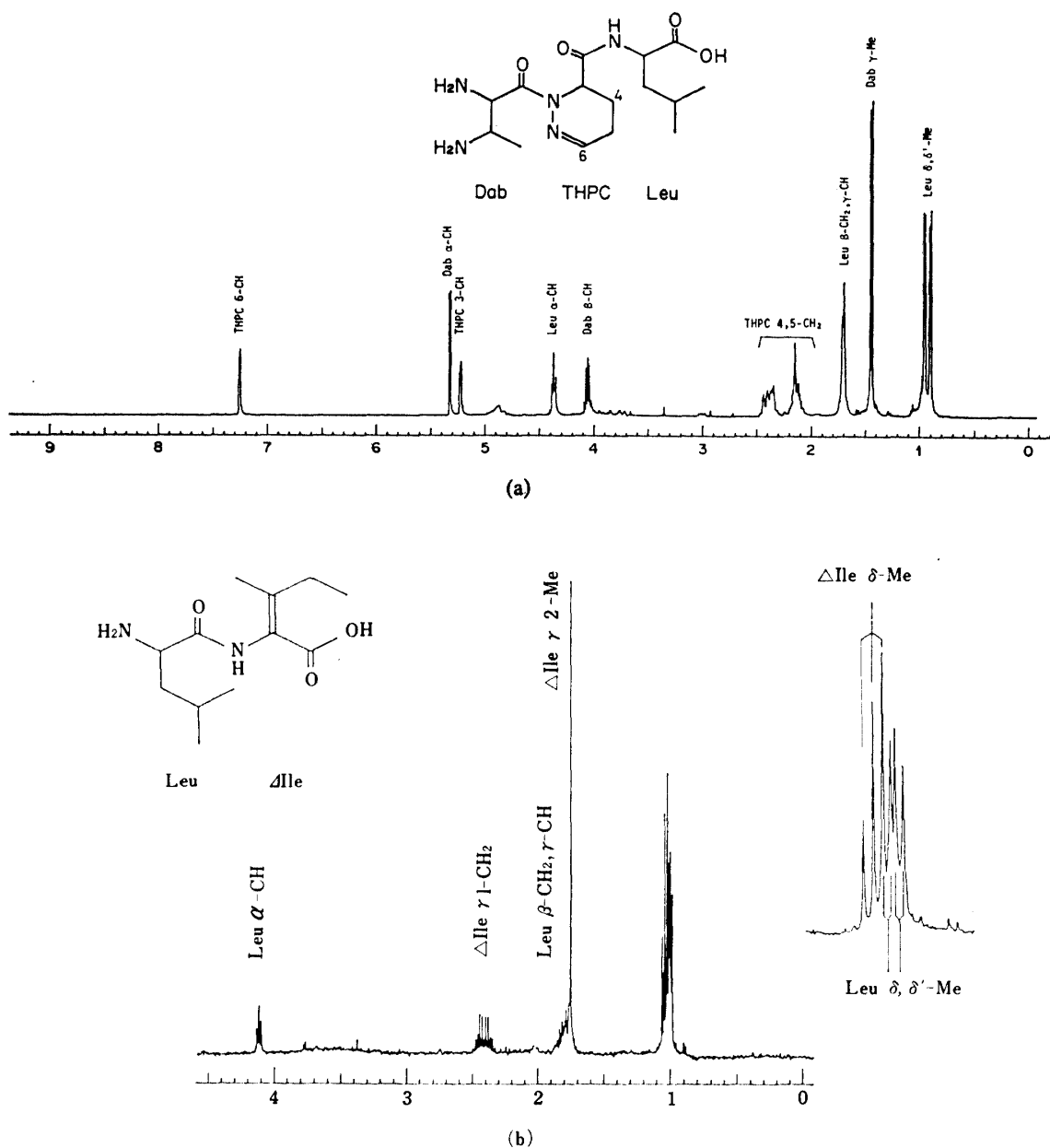
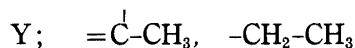
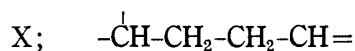


Fig. 4.  $^1\text{H-NMR}$  Spectra of Tripeptide **6** (a) and Dipeptide **7** (b) in  $\text{D}_2\text{O}$ .

In addition to these fragments, Dab was isolated without any racemization and identified to take an *L-erythro* configuration by ORD and  $^1\text{H-NMR}$  spectra ( $J_{\text{H}\alpha-\text{H}\beta} = 8 \text{ Hz}$ ).

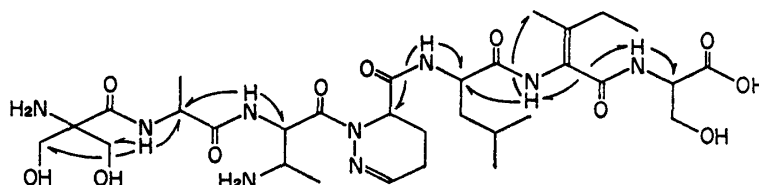
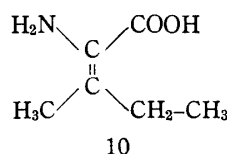
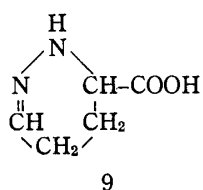
The  $^1\text{H-NMR}$  spectrum of **5** revealed the signals attributed to HMSer ( $\delta$  3.73 ppm, 2 H, 3.92 ppm, 1 H, 3.94 ppm, 1 H) and Ala ( $\delta$  1.39 ppm, 3 H, 4.23 ppm, 1 H), but dansylated **5** could not be obtained, probably due to steric hindrance around the amino group of HMSer. This result indicated the structure of **5** as HMSer-Ala.

From the  $^1\text{H-NMR}$  spectra of **6** and **7**, the following partial structures are suggested for the two unknown amino acids X and Y.



Isolation of these peptide fragments, together with the fact that the  $\alpha$ -amino group of Dab was protected in the parent antibiotic, established the amino acid sequence of **1** as  
 HMSer-Ala-Dab-X-Leu-Y-Ser

The FD mass spectrum of tripeptide **6** exhibited  $(M+H)^+$  and  $(M+Na)^+$  ion peaks at  $m/z$  342 and 364, respectively, and further spectral data allowed us to assign the molecular formula of X as  $C_5H_8N_2O_2$ . Catalytic hydrogenation of **6** over  $PtO_2$  in 1 N AcOH (room temperature, 5 days), followed by acid hydrolysis, liberated Leu, Dab and L-ornithine which were identified by TLC, amino acid analysis and ORD. This result proved the unknown amino acid, X, to be 2,3,4,5-tetrahydropyridazine-3-carboxylic acid (THPC, **9**). Thus the structure of **6** was elucidated as L-erythro Dab-(S)-THPC-L-Leu. Reductive cleavage of the 4-hydroxy-2,3,4,5-tetrahydropyridazine-3-carboxylic acid to  $\beta$ -hydroxy-ornithine has been described.



The EI mass spectrum of an *N*-acetyl derivative of **7** ( $M^+$   $m/z$  284) suggested the molecular formula of  $C_6H_{11}NO_2$  for Y and the  $^1H$ -NMR spectral analysis of **7** revealed the structure of Y as 2,3-didehydroisoleucine ( $\Delta$ Ile, **10**). Hydrogenation of **8** over  $PtO_2$  in 1 N AcOH followed by acid hydrolysis gave Leu, Ser and Ile, confirming the above conclusion and the sequence of **8** as Leu- $\Delta$ Ile-Ser.

In order to determine the stereochemistry of the  $\Delta$ Ile moiety, an NOE experiment was carried out. Upon irradiation of a singlet amide proton of **1** ( $\delta$  9.31 ppm, assignable to the amide proton of  $\Delta$ Ile) in  $DMSO-d_6$ , the negative NOE was observed with singlet methyl protons ( $\delta$  1.65 ppm) of  $\Delta$ Ile indicating its configuration to be *E*. By irradiating four other amide protons separately, negative NOEs were also observed with relevant protons, and the amino acid sequence proposed was again confirmed as shown below.

These experimental results elucidated the linear structure for **1**, having the free amino group of HMSer which could not be dansylated because of its presumable steric hindrance

and Ser as the C-terminal.

Cirratomycin B (**2**) has properties closely related with **1** and its  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra were essentially superimposable with those of **1**, except for the presence of Ala residue instead of Leu. Acid hydrolysis of **2** liberated two moles of L-Ala and one mol of L-Ser along with HMSer and Dab. *N*-Acetylation of **2** also afforded *N*-monoacetyl cirratomycin B and *N*-diacetyl cirratomycin B. Mild acid hydrolysis of **2** gave amino acid segments HMSer-Ala, Dab-THPC-Ala, Ala-Ile and Ala-Ile-Ser and confirmed the structure of cirratomycin B as shown in **2**.

Recently, Dr. Umezawa and his associates have reported independently the structures of antrimycin A-Dv, in which the identity of antrimycin A with cirratomycin B and antrimycin D with cirratomycin A were confirmed, respectively.