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Author	佐藤, 良博(Sato, Yoshihiro) 小田, 泰子(Oda, Taiko) 斉藤, 肇(Saito, Hajime)
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Microbial Transformation of Dehydrogriseofulvin and Griseofulvin : ²H N.m.r. and Mass Spectrometric Studies of Stereochemical Courses of Microbial Hydrogenation and Hydroxylation*

YOSHIHIRO S ATO, T AIKO ODA and HAZIME S AITO (National Cancer Center Research Institute)

佐藤良博,小田泰子,斉藤 肇

We have recently demonstrated that ²H n.m.r. spectroscopy provides very powerful method to study biosynthetic pathways involving hydrogen. ²H N.m.r. chemical shifts, expressed in p.p.m., essentially the same as those of the analogous ¹H isotope. Therefore, ²H n.m.r. signals of griseofulvin and related compounds can be assigned on the basis of known chemical shifts in the corresponding ¹H n.m.r. spectra. We now report on the stereochemical courses of the deuterium atom(s) at C-5' during the microbial transformation of dehydrogriseofulvin (la) and griseofulvin by *Streptomyces cinereocrocatus* NRRL 3443 as studied by ²H n.m.r. and mass spectrometry.

 $(5' - {}^{2}H)$ Dehydrogriseofulvin (1b) (${}^{2}H_{0}$ 73.9, ${}^{2}H_{1}$ 26.1%) was administered to a shaken culture of *S. cinereocrocatus* on the 4th day of the fermentation period. After 3 days, griseofulvin (2a) (${}^{2}H_{0}$ 75.4, ${}^{2}H_{1}$ 24.6%) was isolated from the broth. Since the ${}^{2}H$ n.m.r. resonance of (2a) is at the same position as that of the 5' α -signal of $(5'\alpha,5'\beta-{}^{2}H)$ griseofulvin (2d) (${}^{2}H_{0}$ 21.5, ${}^{2}H_{1}$ 53.6, ${}^{2}H_{2}$ 24.9%) prepared by a previously described method (Figure, A and C), the configuration of deuterium was unequivocally ascribed as 5' α . As shown in the Figure, B, the ${}^{2}H$ n.m.r. spectrum of the mixture of (2a) and (3) (4.2 : 1) exhibits only one signal, corresponding to the 5' α -signal of $(5'\alpha-{}^{2}H)$ griseoflvin (2a). Accordingly, it is concluded that the hydroxylation occurs at the 5' α -position of (2a) without any configurational change of the deuterium, as summarized in the Scheme, A.

The above results were further confirmed by an alternative study of deuteration at the 5'-position by D_2O in the fermentation medium *via* microbial hydrogenation (Scheme, B). A medium containing 50% D_2O and undeuteriated dehydrogriseofulvin (1a) was inoculated with a culture of *S. cinereocrocatus* which had been fermenting for 3 days. The ²H n.m.r spectrum of the purified griseofulvin (2b), which was obtained after 3 days fermentation, is shown in the Figure, D. Because the deuterium atom could be simultaneously incorporated at the 6'-position also under these condition, the deuterium peak, the position of which is in agreement with that of the 5' β -signal, may be a superposition of 5' β - and 6'-signals. However, it was possible to prove that some incorporation of deuterium had occurred at the 6'-position by treatment with neutral alumina (Woelm, activity II) for 48 h, which removes the deuterium at the 5' β -position selectively. The decrease in deuterium content was as

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follows : (2b) ; 0.33 2 H/molecule, 2 H₀ 69.3, 2 H₁ 28.6 and 2 H₂ 2.1%, and (2c) ; 0.13 2 H/molecule, 2 H₀ 86.6 and 2 H₁ 13.4%. In harmony with this, the peak intensity of (2c, 19mg) decreased considerably in comparison with of (2b, 22mg) (Figure, D and E).



FIGURE. ³H N.m.r. spectra of microbial transformation products of dehydrogriseofulvin and griseofulvin, and of $[5'\alpha, 5'\beta^{-3}H]$ griseofulvin, in chloroform (C₆F₆, internal lock) at 15.28 MHz on a JEOL PFT-100/EC-100 spectrometer with proton noisedecoupling.

Treatment of (2c) with neutral alumina for a further 24 h showed that the deuterium content was 0.11 ²H/molecule (²H₀ 89.1, ²H₁ 10.9%). These results indicate that during the course of the microbial hydrogenation, (1a) was transformed to $(5'\beta,6'-{}^{2}H)$ griseofulvin in which deuteriums are incorporated at the 5' β - and 6' α -position in ca 2:1 ratio. Finally, the above results were also supported by mass spectrometric studies of the 5' α -hydroxylation products from $(5'\beta-{}^{2}H)$ - and $(5'\alpha,5'\beta-{}^{2}H)$ -griseofulvin samples.